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CERTIFICATE

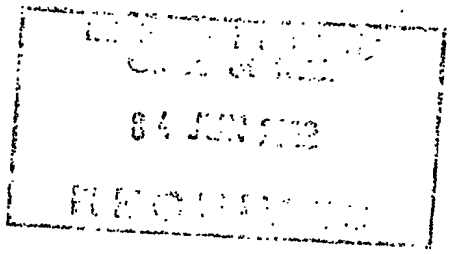
This certificate is issued in support of an application for Patent registration in a country outside New Zealand pursuant to the Patents Act 1953 and the Regulations thereunder.

I hereby certify that annexed is a true copy of the Provisional Specification as filed on 4 June 2002 with an application for Letters Patent number 519371 made by AUCKLAND UNISERVICES LIMITED.

Dated 27 June 2003.

Neville Harris
Commissioner of Patents

519371



Patents Form No. 4

Our Ref: TJ503958

Patents Act 1953

PROVISIONAL SPECIFICATION

IMMUNOMODULATORY CONSTRUCTS AND THEIR USES

We, **AUCKLAND UNISERVICES LIMITED,** a New Zealand company, of Level 10, 70 Symonds Street, Auckland, New Zealand do hereby declare this invention to be described in the following statement:

PT0404503

IMMUNOMODULATORY CONSTRUCTS AND THEIR USES

TECHNICAL FIELD

This invention relates to immunomodulatory constructs and their use. In particular, it relates to constructs which target antigen-presenting-cells for the purpose of enhancing or suppressing a host immune response, and to methods of enhancing antigenicity of compounds.

BACKGROUND ART

Professional antigen-presenting-cells (APC) are essential to initiate a primary immune response in a non-immune, naive animal. The most important APC is the Dendritic Cell (DC), which is found as an interdigitating cell at all regions of the body, at an interface with the environment (i.e. skin and mucosal surfaces such as the lung, airways, nasal passage etc). Antigens presented by DCs are profoundly immunogenic. One important phenotypic marker of the DC is a very high level of surface MHC class II expression. Activated DCs migrate to secondary lymph nodes to "prime" both CD4 and CD8 T cells which proceed as antigen activated effector cells, to proliferate, produce cytokines and regulate the humoral response of B-lymphocytes. Thus, antigen presentation by DC appears to be the obligate first step in any adaptive immune response. Other APCs such as macrophages and B-cells appear to be important in later, secondary responses and by themselves are not effective in the initial priming of a response. Thus the DC is generally regarded as the most important cell to target for enhancement of immune responses.

The targeting of antigens to DC can however be problematic. For example, many peptides by themselves are poorly antigenic and immunogenic because they are not efficiently delivered to APC in vivo. They are equally not taken up by APC very efficiently and do not elicit the second signals required for efficient antigen presentation.

Superantigens are a family of semi-conserved bacterial proteins that target the immune system by binding simultaneously to the T cell Receptor (TcR) via the Vβ domain on T lymphocytes and MHC class II molecules expressed on APC including dendritic cells.

Superantigens (SAgs) are the most potent immune mitogens known and activate large numbers of T cells at femto-attomolar concentrations (10^{-15} - 10^{-18}M).

They cause significant toxicity due to the massive systemic cytokine release by T cells. There are currently 19 members of the staphylococcal and streptococcal superantigen family.

Terman (WO 98/26747) discloses therapeutic compositions employing superantigens. It is suggested that superantigens, in conjunction with one or more additional immunotherapeutic antigens, may be used to either induce a therapeutic immune response directed against a target or to inhibit a disease-causing immune response. Terman further describes the formation of immunotherapeutic antigen-superantigen polymers. Such polymers include those where the superantigen component is coupled to a peptide antigen by a secondary amine linkage. However, there is no teaching or suggestion by Terman that the superantigen component be one from which the TcR binding function has been wholly or partly ablated. Indeed, there is no recognition that a TcR binding is not essential to activation of APCs and to stimulation of an immune response against the antigenic component of the polymer.

Thus, wild-type SAgs, or modified SAgs which retain the ability to bind to TcR, are of little use because they themselves elicit massive, indiscriminate T cell responses by binding to the TcR. This TcR cross-linking appears to be the major cause of their toxicity [12].

There exists a need therefore for improved immunomodulators which exploit the unique features of DC targeting and activation of SAgs to deliver and enhance the T cell recognition of antigens such as peptides that are normally non-immunogenic or have low immunogenicity, yet are efficacious and have low toxicity.

It is an object of the present invention to overcome or ameliorate at least some of the disadvantages of the prior art, or to provide a useful alternative.

SUMMARY OF THE INVENTION

According to a first aspect there is provided an immunomodulator which comprises an antigen-presenting- cell (APC) targeting molecule coupled to an immunomodulatory antigen, wherein said APC-targeting molecule mimics a superantigen but does not include a fully functional T-cell receptor binding site.

According to a second aspect there is provided an immunomodulator which comprises an antigen-presenting cell (APC) targeting molecule coupled to an

immunomodulatory antigen, wherein said APC-targeting molecule is a molecule which is structurally a superantigen but for a disrupted T-cell receptor binding site such that the molecule has little or no ability to activate T-cells.

Preferably the T-cell receptor binding site, or at least part thereof, of the antigen-presenting- cell (APC) targeting molecule is derived from *Staphylococcus aureus* and/or *Streptococcus pyogenes*. Particularly preferred is a targeting molecule derived from SPE-C and the preferred truncation involves deletion of residues 22-90 from the wild-type SPE-C sequence. However it will be clear to those skilled in the art that other SAgs which have a similar or otherwise known TcR binding region of the molecule may also be advantageously used, for example SMEZ, SEA and the like.

The T-cell receptor binding site, or at least a part thereof, of the antigen-presenting- cell (APC) targeting molecule can also been modified by substitution or addition, to remove or minimise TcR binding. An example of such a targeting molecule is SPEC-Y15A R181Q of the present invention.

A particularly preferred intermediate in the generation of the immunomodulator is Y15A.C27S.N79C.

Preferably the coupling between the antigen-presenting- cell (APC) targeting molecule and the immunomodulatory antigen will be reversible. However, it will be understood from the following description that what is preferably required is that the antigen-presenting- cell (APC) targeting molecule is capable of releasing the immunomodulatory antigen so that it is correctly presented by the APC. Thus, it would also be clear that the release of the immunomodulatory antigen from the immunomodulator may be achieved by intracellular or intralysosomal enzymatic cleavage. This process may be assisted by introducing the appropriate proteolytic site into the coupling region of the immunomodulator. The release may also be achieved by chemical means, which includes redox reactions involving disulphides and free sulphydryl groups. This process may also be assisted by introducing into the coupling region certain amino acid residues, eg. cysteine.

Preferably the immunomodulatory antigen is a protein, a polypeptide and/or a peptide however similar principles may be applied to antigens which are non-proteinaceous, for example nucleic acids or carbohydrates.

The immunomodulatory antigen may be entirely non-immunogenic when not coupled to the antigen-presenting cell (APC) targeting molecule but the immunomodulators of the present invention may also incorporate antigens which are immunogenic, in order to improve their efficacy. Thus the present invention is equally applicable to for example to new vaccines as it is to those which are already known and used but which can be improved by means of the immunomodulators of the present invention.

According to a third aspect there is provided a pharmaceutical composition comprising an immunomodulator according to the present invention and a pharmaceutically acceptable carrier, adjuvant, excipient and/or solvent.

According to a fourth aspect there is provided a vaccine comprising an immunomodulator according to the present invention.

According to a fifth aspect there is provided a method of therapeutic or prophylactic treatment of a disorder which requires the induction or stimulation of the immune system, comprising the administration to a subject requiring such treatment of an immunomodulator or of a pharmaceutical composition according to the present invention.

Preferably the disorder is selected from the group consisting of bacterial, viral, fungal or parasitic infection, autoimmunity, allergy and/or pre-neoplastic or neoplastic transformation.

According to a fifth aspect there is provided the use of an immunomodulator according to the first or the second aspect for the preparation of a medicament for the therapeutic or prophylactic treatment of a disorder which requires the induction or stimulation of the immune system.

The preferred disorder is selected from the group consisting of bacterial, viral, fungal or parasitic infection, autoimmunity, allergy and/or pre-neoplastic or neoplastic transformation.

According to a sixth aspect there is provided a method of preparing an immunomodulator comprising the steps of:

a introducing a modification and/or a deletion into the T-cell binding site of an antigen-presenting cell (APC) targeting molecule which is structurally a superantigen, and

b coupling thereto and immunomodulatory antigen.

Preferably the antigen-presenting cell (APC) targeting molecule is selected from the group of SPE-C, SMEZ and SEA and more preferred are the antigen-presenting cell (APC) targeting molecules SPE-C Y15A. R181Q or SPEC (-20-90). Even more preferred is SPEC-Y15A.C27S.N79C.R181Q

It will be understood however that more than one antigen-presenting cell (APC) targeting molecule may be employed and that a combination of immunomodulators may be used in any treatment.

BRIEF DESCRIPTION OF THE FIGURES

Figure 1. Antigenicity of SAG:PCC conjugate

Figure 2. Immunogenicity of SPEC:PCC conjugate

Figure 3. Proliferation of 5C.C7 LN cells to SPEC-CytC vs MHC-/-SPEC-CytC and free CytC peptide in vitro

Figure 4. Proliferative responses of SMEZ TcR mutants (Figures A and B represent similar data from different experiments)

Figure 5. Proliferative responses of 5C.C7 LN Cells with PCC-SAg Complexes. (Legend: The red line indicates the proliferative response to PCC protein alone. The blue square line shows that the response to PCC-SPEC is 100-fold more antigenic than the unconjugated PCC protein. The green square line is the response to PCC-SMEZ and is approximately 80 fold more antigenic than to unconjugated PCC protein. The black square shows the response to PCC conjugated to SPEC defective in MHC class II binding is no greater than the response to unconjugated PCC protein. The triangles represent the proliferative response of T cells to SAG and PCC together as a mixture but not conjugated).

Figure 6. Effect of SAG-LCMV peptide coupling on antigenicity for CTL.

Figure 7. Inhibition of tumour growth by immunisation with SMEZ-LCMV complexes (Figures A and B represent similar data from different experiments).

DESCRIPTION OF THE PREFERRED EMBODIMENT

The present invention is based at least in part on an unexpected observation that a molecule which mimics a superantigen but which lacks a fully functional TcR binding site can, when coupled to an immunomodulatory antigen, bind and activate APCs to a degree not previously known or suspected. Thus, such immunomodulatory constructs are effective in antigen presentation without the requirement to bind to the TcR. This is of particular relevance to moieties which have low or nonexistent immunogenicity, such as peptides, proteins, nucleic acids, whole viruses etc

The applications of this technology rely on the ability to generate a wide variety of immunomodulatory reagents that combine the delivery capacity and APC activating potential of the TcR ablated superantigens with the specificity of a coupled antigen.

A preferred use of this technique is to enhance responses to synthetic peptides as has been displayed herein with the PCC peptide. However, the antigen need not be a synthetic peptide, but could be a native or recombinant polypeptide, protein of even whole disabled virus. Further, the antigen need not be proteinaceous and may be a nucleic acid or carbohydrate antigen. Also, the present invention can be applied to antigens which are immunogenic, by improving immunogenicity or reducing the quantity of antigen required to induce an immune response

Peptides can be designed to be either stimulatory (i.e. generate agonist responses) or immunosuppressive (i.e. generate antagonist responses) to induce tolerance depending on the primary sequence of the peptide. This is useful in either promoting immunity for vaccination against pathogens such as viruses, bacteria and other micro-organisms, or for generating specific anti-tumour immunity using tumour specific peptides.

Antagonist responses induce T cell tolerance to antigen and might be useful to suppressing unwanted autoimmune reaction to self-antigens eg. proteins and/or nucleic acids, in the case of diseases such as multiple sclerosis, diabetes or rheumatoid arthritis.

Many autoimmune diseases have their basis in an auto-reactive T cell response to self antigens. Diseases such as rheumatoid arthritis, multiple sclerosis and diabetes mellitus are such examples.

The present invention will now be exemplified more particularly with reference to non-limiting examples.

EXAMPLES

Example 1: Cloning and expression of superantigen genes

Genes coding for individual wild-type superantigens were isolated and cloned directly from the DNA of isolates of *Staphylococcus aureus* or *Streptococcus pyogenes* using polymerase chain reaction (PCR) and oligonucleotides inferred from published sequences. All wild type sequences have been confirmed by DNA sequencing.

The methods used for isolation, cloning and sequencing are standard laboratory procedures and are described in for example Goshorn SC, Schlievert PM. 1988. Nucleotide sequence of streptococcal pyrogenic exotoxin type C. Infect Immun. 56(9):2518-20. Proft T, Moffatt SL, Berkahn CJ, Fraser JD. 1999. Identification and characterization of novel superantigens from Streptococcus pyogenes. J Exp Med. Jan 4;189(1):89-102, all incorporated herein by reference.

A summary of the SPE-C single domain molecule and its derivation is set out below, including the comparative proliferative response of human T cells.

C-terminal Single Domain of SPE-C

(C-terminal single domain references the term "truncated SPE-C" and is a reference to the explicitly stated SPEC-(-20-90). The parenthesized numbers represent that part of the native SPE-C that has been deleted as outlined in the procedure below)

Vector: pGEX-3C (variation of pGEX-2T)
Host: DH5□
Antibiotic resistance: Ampicillin
Restriction sites: 5' BamH1, 3' EcoR1
Brief Expression protocol:

*. Grow overnight culture in LB-Amp at 37°C with shaking.

*. Dilute overnight culture 1:10 with pre-warmed LB-Amp.

*. Grow for another hour or until the absorbance at 600nm is 0.9.

*. Cool culture to 30°C.

*. Induce protein expression with 0.1mM IPTG.

*. Incubate at 30°C with shaking for 4 – 5 hours.

*. Harvest cells and resuspend in 10 mls of GSH Buffer 1 (25 mM Tris.Cl pH 7.4 / 50 mM NaCl / 1 mM EDTA) for every 1 gram of pellet.

*. Sonicate to lyse cells and release soluble fusion protein.

*. Spin lysate to remove insoluble material.

*. Dialyse lysate overnight in GSH Buffer 1 to remove endogenous GSH (this step will increase yields but is not essential).

*. Purify GST-Fusion protein from bacterial proteins using GSH agarose affinity chromatography.

*. Cut purified fusion protein overnight with 3C protease at 4°C (NB to add DTT)

*. Dialyse cut fusion protein into 10 mM PO_4 pH 6.0 overnight.

*. Purify C-terminal Single Domain from GST using cation exchange chromatography (ie MonoS column – elute with pH gradient 6.0 – 7.0 over 20 column volumes)

Sequence details:

Includes residues 1 – 21 of SPE-C, 4 amino acid linker which is the Factor X protease cleavage site, and then residues 91 – 208 of SPE-C.

DNA sequence (Factor X sequence shown in gray):

```
GAC TCT AAG AAA GAC ATT TCG AAT GTT AAA AGT GAT TTA CTT TGC GCA TAC
ACT ATA ACT CCT ATC GAA GGT CGT ACG CCT GCT CAA AAT AAT AAA GTA AAT
CAT AAA TTA TTG GGA AAT CTA TTT ATT TCG GGA GAA TCT CAA CAG AAC TTA
AAT AAC AAG ATT ATT CTA GAA AAG GAT ACC GTA ACT TTC CAG GAA ATT GAC
TTT AAA ATC AGA AAA TAC CTT ATG GAT AAT TAT AAA ATT TAT GAC GCT ACT
TCT CCT TAT GTA AGC GGC AGA ATC GAA ATT GGC ACA AAA GAT GGA AAA CAT
GAG CAA ATA GAC TTA TTT GAC TCA CCA AAT GAA GGG ACT AGA TCA GAT ATT
TTT GCA AAA TAT AAA GAT AAT AGA ATT ATC AAT ATG AAG AAC TTT AGT CAT
TTC GAT ATT TAT CTT GAA AAA TAA
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Protein Parameters:

Protein Sequence:

```
DSKKDISNVKSDLLCAYTITPIEGRTPAQNNKVNHKLLGNLFISG
ESQQNLNNKIILEKDTVTFQEIDFKIRKYLMDNYKIYDATSPYVS
GRIEIGTKDGKHEQIDLFDSPNEGTRSDIFAKYKDNRIINMKNFSH
FDIYLEK Stop
```

Molecular Weight: 16543

Theoretical pI: 7.02

Theoretical Extinction data (6M Guanidine-HCl/20mM phosphate, pH 6.5)

Assuming all cysteines are reduced:

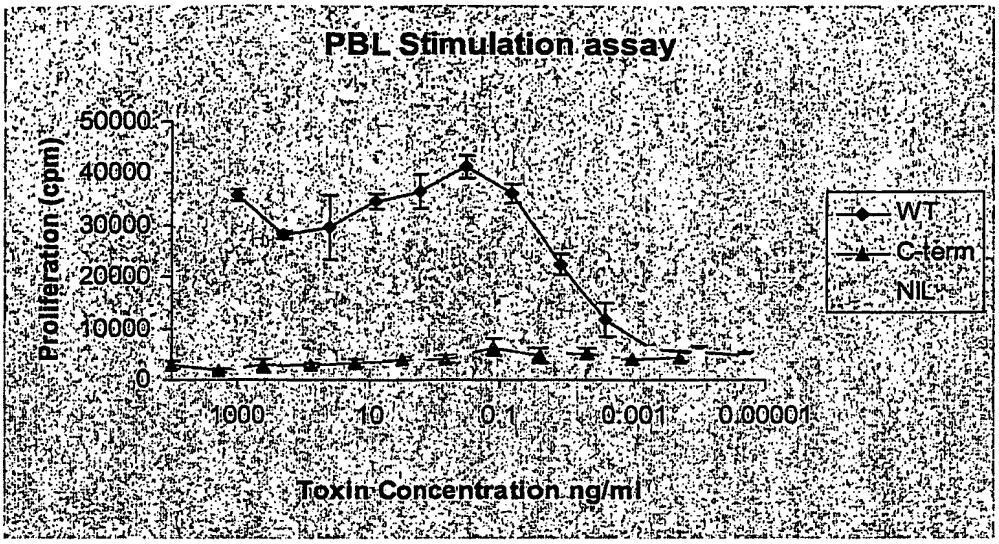
Molar A280: 8960

A280/cm (1mg/ml): 0.542

Activity of C-terminal Domain SPE-C

PBL stimulation assay:

Peripheral blood lymphocytes are isolated from blood using Hypaque-Ficoll. A 5 fold serial dilution of toxin in RPMI (complete) is set up in a 96 well plate. 1 x 10^5 PBLs is added to each well containing varying concentrations of toxins. The plates are left to incubate for 3 days after which time [^{3}H]Thymidine is added to each well to measure proliferation. The cells are harvested the next day and [^{3}H]Thymidine incorporation is measured.



The above figure shows that the C-terminal domain SPE-C does not have stimulatory activity above background with human PBLs. This is most likely due to the fact that it cannot interact with the TcR on T cells or cross-link MHC on the cell surface of antigen presenting cells.

Example 2: Ablation of TcR binding residues in superantigens

The gene from SPE-C was derived from a patient isolate of Streptococcus pyogenes by PCR using synthetic primers to the 5' and 3' end of the genes. These primer sequences were obtained from the published sequence of Goshorn SC, Schlievert PM. 1988. Nucleotide sequence of streptococcal pyrogenic exotoxin type C. Infect Immun. 56(9):2518-20. GenBank accession number M35514. Any other Streptococcus pyogenese isolate can also be used for this purpose.

Primers used to amplify the SPEC gene are listed in table 1 as SPEC-N-terminal and SPEC-C-terminal. The sequence was confirmed by DNA sequencing.

The full length SPE-C gene was sub-cloned into the expression vector pGEX-3T (Pharmacia) following manufacturers instructions which was used to transform the bacteria *E. coli* using standard procedures (Maniatis et al,). Recombinant SPE-C fused to glutathione-S-transferase was purified from *E. coli* cultures using Glutathione Agarose affinity chromatography.

Table 1: Primers used for amplification of the SPEC gene and introduction of mutations or truncations

SPEC – N-terminal	CGGGATCCGACTCTCAAGAAAGACA	
SPEC – C-terminal	CTGAATTCTTATTTTTCAAGAT	
SPEC- Y15A	GATTTACTTTGTGCATACAC	GTGTATGCACAAAGTAAATC
SPEC- N79C	ATATTCTTTGTTCTCACA	TATAAGAAACAAGAGTGT
SPEC- Y15C	GATTTACTTTGTGCATACAC	GTGTATGCACAAAGTAAATC
SPEC- R181Q	GAAGGGACTCAATCAGATATTTTTGC	GACAAAATATCTGATTGAGTCCCTTC
SPEC-(-20-90)	ATCGAAGGTCGTACGCCTGCTCAAAATAATAAAG	ACGACCTTCGATAGGAGTTATAGTGTAT
SPEC- C27S	GATTATAAAGATTCCAGGGTAA	TTACCCTGGAATCTTTATAATC

Sequential introduction of current mutations into SPE-C.

1. SPEC – C27S
To remove a naturally occurring cysteine that interferes with the coupling of antigen to the preferred site at N79C.

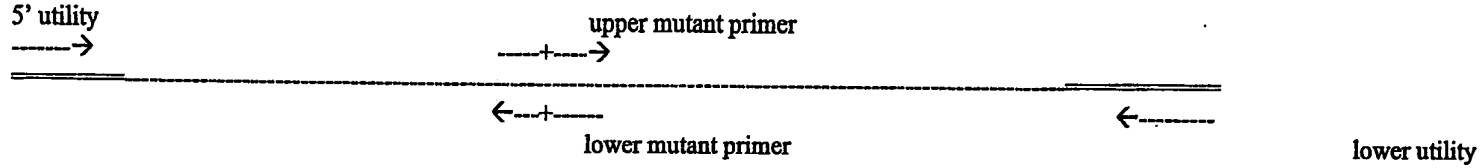
2. SPEC – C27S, N79C
To introduce the coupling point for antigen. This position was chosen from the crystal structure of SPE-C to be well exposed and to not interfere with MHC class II binding.

3. SPEC – C27S, N79C, Y15A
To destroy TcR binding

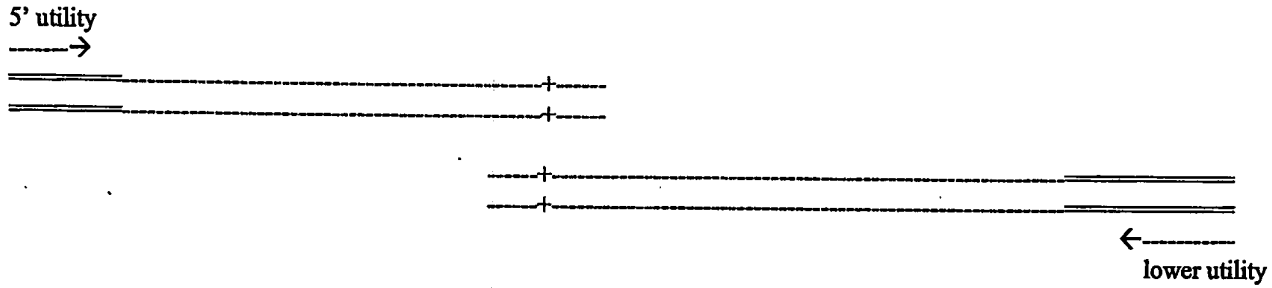
4. SPEC – C27S, N79C, R181Q
To further limit binding of T cell Receptors.

PCR overlap

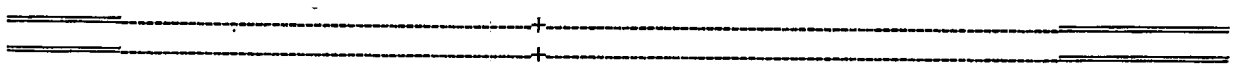
1st round - amplification in separate tubes preoduces two overlapping products.
(+ indicates the position of the mutation to be introduced.
==== represents vector sequence
..... Represents target sequence)



2nd round - combines the products of the first amplifications and amplifies with the utility primers



Final product



Product is subcloned into the expression vector.

Glutathione-Agarose was manufactured according to previously published methods[22,23]. Recombinant SPE-C protein was purified after cleavage of the fusion protein with trypsin by ion cation exchange chromatography according to the method described in reference 5 which is incorporated herein. Purified SPE-C was crystalised and the 3-D structure determined according to Roussel, 1997 (Ref 26), which is incorporated herein by reference.

Identification of amino acids in SPE-C that are important to TcR binding were determined by a combination of molecular modelling of the 3D crystal structure of SPE-C and comparison with known TcR binding residues of the related superantigen SEB.

Rational mutagenesis of residues thought to be part of the TcR interface.

TcR binding residues were targeted by site-directed mutagenesis using the method of PCR overlap [24].. The synthetic primers used to produce each mutation are described in the accompanying table of primers (Table 1). The process of introducing two mutations was performed sequentially as described in the accompanying diagrams describing the sequential introduction of successive mutations in SPE-C and the method of PCR overlap which is used to introduce said mutations.

The mutant form of SPE-C of the present invention was confirmed by automated DNA sequencing (Licor Inc. USA) then inserted into the pGEX expression vector between the BamH1 and EcoR1 restrictions sites according to the manufacturers description of the cloning site for this vector. A strain of *E. coli* DH5a was transformed with the recombinant vector and colonies expressing the pGEX fusion protein were isolated to grow up in large scale cultures for the purposes of protein purification.

To test the effects of mutations in the TcR binding site, recombinant proteins were added to cultures of human peripheral blood lymphocytes, isolated by standard techniques (for examples of techniques see Handbook Of Experimental Immunology, ed. D.M. Weir, Blackwell Scientific Publications), to determine what concentration of recombinant SPE-C was required to stimulate the proliferation of human T cells. Wild-type SPE-C normally stimulates human T cells at 50% of maximal proliferation at 0.2 pg/ml. Two residues were identified from

these studies that when mutated, reduce T cell proliferation by 1,000,000 fold when compared to wild-type SPE-C. These residues are Y15 and R181. SPE-C molecules with these two mutations (SPEC-Y15A, R181Q) no longer stimulate human T cells

Amino acid residues in superantigens that are important to the interaction with T cell Receptor have been identified from the present mutational studies and those of others (Table 2 below). Loss of T cell activation is determined by *in vitro* T cell proliferation assays (see below) and compared to the activity of wild-type molecule. All mutants are also assessed for their ability to bind to MHC class II by a number of assays including direct binding to MHC class II expressing B cells as well as Biacore studies with soluble forms of both superantigen mutant and MHC class II.

3D crystal structures of the superantigens SEC3 bound to a murine T cell Receptor [4,13] provides the most complete information about the nature of superantigen/TcR interaction but is limited to those with SEC3-like activity. Most single point mutations result in only a small loss in superantigen activity due to only small reductions in binding affinity to the TcR. It is rare to find a single mutation that completely abrogates all mitogenic potential. Only SPE-C Y15A has been shown (Yamoaka et al, Infect. Immunol. 1998 66:5020 and McCormick et al, J. Immunol. 2000 165: 2306-2312) to cause more than a 1000-fold reduction in T cell responses to a superantigen.

The combined mutations producing SPE-C Y15A, R181Q of the present invention generates a form of SPE-C that has no detectable T cell activating potential.

By homology modelling of the 3D crystal structures of other SAgs important regions for binding to the TcR can be identified and corresponding mutants prepared and used to generate immunomodulators of the present invention.

Example 3: T cell proliferation assay The T cell proliferation assay used was a standard technique described for example in Hudson KR et al, incorporated herein by reference

Purified recombinant mutant superantigens are incubated with freshly isolated human peripheral blood lymphocytes at varying dilutions in microtitre

plates for 3 days. A fixed amount of ^3H thymidine is added on the 3rd day and the cells are harvested on day 4. The amount of ^3H thymidine incorporated into the cellular DNA is measured by scintillation autography and is a direct measure of the degree of cell proliferation. Mutant superantigens are compared to wild-type superantigens. The proliferative potential of a given superantigen or mutant is expressed as the concentration required to induce 50% of its maximal stimulation ($P_{50\%}$).

A fully ablated TcR binding negative superantigen is defined herein as one that displays less than about 0.0001% of proliferative activity of the wild-type superantigen (i.e. a 1 million-fold reduction in activity).

Table 2. Amino acid residues implicated in TcR binding of known superantigens.

	Residues implicated in TcR binding sites	References
SEA	N25, P206, D207	5,14
SEB	N23, Y90	12
SEC3	G19, T20, N23, Y26, N60, Y90, V91, G102, K103, V104, G106, F176, Q210	13,4
SEE	N23, S206, N207	5,14
TSST	Tyr115, Glu132, His135, Ile140, His141 and Tyr144, Q136A	15,16
SPE-C	Y15*, R181*	Present invention
SMEZ-2	D42N, W75L,Y77A, K182Q, S7A, N11A, D181A	Present invention

Those in bold indicate mutations that decrease activity by more than 100-fold.

* Mutation that totally ablates T cell responses

Primary DNA sequences of the wild-type and the mutant form of SPE-C are detailed below:

SPE-C wild type (from GenBank)

Streptococcus pyogenes pyrogenic exotoxin C gene, 5' end cds

```
GACTCTAAGA AAGACATTTC GAATGTTAAA AGTGATTTAC TTTATGCATA CACTATAACT
CCTTATGATT ATAAAGATTG CAGGGTAAAT TTTTCAACGA CACACACATT AAACATTGAT
ACTCAAAAAT ATAGAGGGAA AGACTATTAT ATTAGTTCCG AAATGTCTTA TGAGGCCTCT
```

CAAAAATTTA AACGAGATGA TCATGTAGAT GTTTTTGGAT TATTTTATAT TCTTAATTCT
CACACCGGTG AGTACATCTA TGGAGGAATT ACGCCTGCTC AAAATAATAA AGTAAATCAT
AAATTATTGG GAAATCTATT TATTTCGGGA GAATCTCAAC AGAACTTAAA TAACAAGATT
ATTCTAGAAA AGGATATCGT AACTTTCCAG GAAATTGACT TTAAAATCAG AAAATACCTT
ATGGATAATT ATAAAATTTA TGACGCTACT TCTCCTTATG TAAGCGGCAG AATCGAAATT
GGCACAAAAG ATGGGAAACA TGAGCAAATA GACTTATTTG ACTCACCAAA TGAAGGGACT
AGATCAGATA TTTTTGCAAA ATATAAAGAT AATAGAATTA TCAATATGAA GAACTTTAGT
CATTTCGATA TTTATCTTGA A

Protein Sequence – wild type

DSKKDISNVK SDLLYAYTIT PYDYKDCRVN FSTTHTLNID TQKYRGKDYY ISSEMSYEAS
QKFKRDDHVD VFGLFYILNS HTGEYIYGGI TPAQNNKVNH KLLGNLFISG ESQQNLNNKI
ILEKDIVTFQ EIDFKIRKYL MDNYKIYDAT SPYVSGRIEI GTKDGKHEQI DLFDSPNEGT
RSDIFAKYKD NRIINMKNFS HFDIYLE

SPEC- Y15A.C27S.N79C.R181Q

GACTCTAAGA AAGACATTTC GAATGTTAAA AGTGATTTACT TTATGCATA CACTATAACT
GATTTACT TTGTGCATA CAC
C27S
CCTTATGATT ATAAAGATTG CAGGGTAAAT TTTTCAACGAC ACACACATT AAACATTGAT
GATT ATAAAGATTC CAGGGTAA
ACTCAAAAAT ATAGAGGGAA AGACTATTAT ATTAGTTCCGA AATGTCTTA TGAGGCCTCT
N79C
CAAAAATTTA AACGAGATGA TCATGTAGAT GTTTTTGGATT ATTTTATAT TCTTAATTCT
ATAT TCTTTGTTCT
CACACCGGTG AGTACATCTA TGGAGGAATT ACGCCTGCTCA AAATAATAA AGTAAATCAT
CA
AAATTATTGG GAAATCTATT TATTTCGGGA GAATCTCAACA GAACTTAAA TAACAAAATT

ATTCTAGAAA AAGATATCGT AACTTTCCAG GAAATTGACT TTAAAATCAG AAAATACCTT

ATGGATAATT ATAAAATTTA TGACGCTACT TCTCCTTATG TAAGCGGCAG AATCGAAATT

GGCACAAAAG ATGGGAAACA TGAGCAAATA GACTTATTTG ACTCACCAAA TGAAGGGACT
GAAGGGACT
R181Q
AGATCAGATA TTTTTGCAAA ATATAAAGAT AATAGAATTA TCAATATGAA GAACTTTAGT
CAATCAGATA TTTTTGC

CATTTCGATA TTTATCTTGAA

Protein Sequence (combined mutants)

DSKKDISNVK SDLLAAYTIT PYDYKDSRVN FSTTHTLNID TQKYRGKDYY ISSEMSYEAS
QKFKRDDHVD VFGLFYILCS HTGEYIYGGI TPAQNNKVNH KLLGNLFISG ESQQNLNNKI
ILEKDIVTFQ EIDFKIRKYL MDNYKIYDAT SPYVSGRIEI GTKDGKHEQI DLFDSPNEGT
QSDIFAKYKD NRIINMKNFS HFDIYLE

Example 4: Purification of recombinant wild-type and mutant proteins

Recombinant wild-type or mutant superantigens are expressed in *E. coli*. Two commercial vectors pGEX-2T (Pharmacia) and pET32A (New England Biolab) have been modified to introduce a new proteolytic cleavage site between the fusion protein and the superantigen. Separation of the two halves of the fusion protein is accomplished with the highly specific 3C protease that only cleaves at the single recognition site.

Two methods are currently used to purify fusion proteins.

a. pGEX-2T produces a fusion protein with the N-terminal component as the Glutathione S-Transferase linked to the superantigen sequence through a protein linker that contains a 3C-protease cleavage site. The fusion protein is purified from the crude bacterial lysate in single step purification on glutathione agarose. Fusion protein is eluted from the glutathione agarose with a buffer containing 5mM glutathione and cleaved by the addition of recombinant 3C protease. Superantigen is further purified by ion exchange HPLC chromatography.

b. pET32-A-3C. Protein is expressed as a stable thioredoxin fusion protein with a 6 histidine tag allowing single-step purification by metal chelation chromatography. Separation of the thioredoxin from superantigen is achieved by cleavage with recombinant 3C protease followed by HPLC ion exchange chromatography.

Expression and purification of the recombinant protein

E.coli transformants are grown overnight at 37°C in a small 100 ml starter culture of Luria Broth (LB) containing 50 mg/ml ampicillin. A 1 litre culture is seeded in the morning and grown to mid-log phase, when IPTG is added to 0.1 mM to induce expression of the fusion protein. The culture is continued for 3 hours at which time cells are pelleted by centrifugation and disrupted by a combination of lysozyme and sonication. The clarified lysate is passed over either

a 5 ml GSH agarose column or a Ni-NTA column. After thorough washing, bound protein is eluted by either 5 mM GSH (GSH agarose) or a buffer containing imidazole (MC chromatography).

The fusion protein is cleaved overnight at room temperature by recombinant 3C protease at a ratio of 1:500 (i.e. 2 mg 3C protease to 1 mg fusion protein). Superantigen is separated from fusion protein by two rounds of cation exchange chromatography. Protein is filter sterilised and stored at 1 mg/ml at 4°C until required.

Introduction of disulphide coupling sites into SPE-C

An exposed cysteine residue has been introduced into the N-terminus of a TcR negative SPE-C at position N79. N79 is located within the putative TcR binding site. Several positions were tested before a residue was identified that met the following criteria

a. Surface exposed and accessible
b. Displayed efficient coupling of synthetic peptide
c. Did not interfere with MHC class II binding
d. Did not render the resulting SAG:peptide conjugate insoluble.

In addition to the introduced cysteine, a naturally occurring cysteine residue at position 27 was mutated to serine to avoid complications with refolding and interference with coupling.

The mutant of SPE-C used herein to provide examples of in vitro and *in vivo* immunomodluatory activity is SPEC-Y15A.C27S.N79C.R181Q, which is a composite of all mutations so far described above that abrogates TcR binding (Y15A and R181Q), introduce an efficient coupling residue (N79C) and removes a naturally occuring cysteine which interfered with coupling (C27S)

Example 5: A truncated version SPEC lacking the N-terminal domain

In addition to the SPEC- SPEC-Y15A.C27S.N79C, an SPEC truncated mutant has been developed by deleting residues 22-90 (SPEC(-20-90))from the wild-type sequence This removes the entire TcR binding region plus the small N-terminal domain. This truncated mutant expresses very well in *E. coli*, is soluble and retains MHC class II binding activity. A cysteine residue has beenintroduced at position 92 to effect antigen coupling using the same method as described for the full length SPEC-Y15A.C27S.N79C molecule. The importance of this mutant

is that it is much smaller, less antigenic (less likely to promote anti-SPEC antibody responses), and will be entirely devoid of any TcR binding ability. It is most unlikely that this truncated SPEC will have any toxicity effects *in vivo that are normally associated with wild-type toxins.*

The primary nucleotide sequence of truncated version of SPE-C is detailed below:

DNA sequence (Factor X sequence shown in gray):

```
GAC TCT AAG AAA GAC ATT TCG AAT GTT AAA AGT GAT TTA CTT TGC GCA TAC ACT
ATA ACT CCT ATC GAA GGT CGT ACG CCT GCT CAA AAT AAT AAA GTA AAT CAT AAA
TTA TTG GGA AAT CTA TTT ATT TCG GGA GAA TCT CAA CAG AAC TTA AAT AAC AAG
ATT ATT CTA GAA AAG GAT ACC GTA ACT TTC CAG GAA ATT GAC TTT AAA ATC AGA
AAA TAC CTT ATG GAT AAT TAT AAA ATT TAT GAC GCT ACT TCT CCT TAT GTA AGC
GGC AGA ATC GAA ATT GGC ACA AAA GAT GGA AAA CAT GAG CAA ATA GAC TTA TTT
GAC TCA CCA AAT GAA GGG ACT AGA TCA GAT ATT TTT GCA AAA TAT AAA GAT AAT
AGA ATT ATC AAT ATG AAG AAC TTT AGT CAT TTC GAT ATT TAT CTT GAA AAA TAA

Protein Sequence
D S K K D I S N V K S D L L C A Y T I T P I E G R T P A Q N N K V N H K L
L G N L F I S G E S Q Q N L N N K I I L E K D T V T F Q E I D F K I R K Y
L M D N Y K I Y D A T S P Y V S G R I E I G T K D G K H E Q I D L F D S P
N E G T R S D I F A K Y K D N R I I N M K N F S H F D I Y L E K Stop
```

Example 6: TcR binding defective versions of SMEZ and SEA

In addition to SPE-C, TcR binding mutants of both SMEZ and SEA using site directed mutagenesis have been prepared. Comparative data of mutant vs wild-types on T cell proliferation is presented in table 3.

Table 3. SMEZ mutants defective in TcR binding

Mutant	P50% (pg/ml)	Reduction
SMEZ-2 wild type	2.0 pg/ml	
SMEZ-2 W75L	>10ng/ml	>100,000
SMEZ-2 D42N	10ng/ml	10,000
SMEZ-2 W75L.D42N.K182Q	>10ng/ml	>100,000
SMEZ-2 Y18A	>10ng/ml	>100,000
SMEZ-2 W75L.D42N.K182Q. Y18A.	>10ng/ml	>100,000

The aim was to produce mutants which stimulate T cells at, for example, about 0.0001% of the activity of the wild type SAG. In addition, a cysteine residues is introduced in the same position relative to N79 in SPE-C.

Including two other superantigens is important to determine whether enhancement of immunogenicity is a feature of all superantigens, or specific to SPE-C. It is clearly broadly applicable, using the principles and techniques described herein.

Similar truncation mutants can be made for other superantigens such as SEA and SMEZ, using the methodology employed for the SPE-C mutants and the information on the Tcell receptor binding regions of the SAGs already published (for example reference #4, incorporated herein by reference).

Example 7: Peptide coupling procedure

Both protein and peptide are stored in 10 mM phosphate pH6.0 under nitrogen to prevent oxidation and auto-dimerisation through the free cysteine.

Synthetic peptide containing a C-terminal cysteine residue and SPEC-Y15A.C27S.N79C are mixed together and incubated at room temperature for 1 hour at a molar ratio of 1:2 in a alkaline buffer containing 1 μM Cu^{2+}. The copper acts as a redox catalyst. In the example below, a synthetic peptide of the pigeon cytochrome C (PCC) is provided, but this method will work for other peptides also so long as a free sulphur atom is present in the peptide.

SPEC-Y15A.C27S.N79C.R181Q (MW 26,500) 10 mg/ml (380 mM)	PCC peptide (RADLIAYLKQATKC) (MW 1400) 10 mg/ml (700 mM)	Buffer
100 μl	10 μl	200mM Tris pH8.0, 1 μM $CuSO_4$

Routinely >80% of SPEC-Y15A.C27S.N79C.R181 is shown to couple to peptide in a ratio of 1:1 Efficiency of coupling is assessed by SDS polyacrylamide gel electrophoresis. The SPEC-Y15A.C27S.N79C:peptide conjugate has a slower mobility on SDS PAGE consistent with an increase in molecular weight from the addition of a single peptide. Addition of 1mM dithiothreitol (DTT) to the conjugate prior to SDS PAGE increases the electrophoretic mobility consistent with a reduction in molecular weight . This indicates that peptide coupling is via a reversible disulphide bond formation - a feature deemed important for dissociation of peptide once inside the APC.

Example 8: Testing of responses to SAG:peptide conjugates

The 5C.C7 T cell Receptor transgenic mouse

This mouse was obtained from The Malaghan Institute for Medical Research, Wellington School of Medicine, Mein St Wellington South, New Zealand

These mice were first generated by Berg et al (Ref 17).

The 5C.C7 transgenic mouse was originally constructed by Berg et al. [17]. This mouse is transgenic for a TcR specific for the pigeon cytochrome C (PCC) peptide presented by mouse I-A^d. Greater than 80% of mature T cells from 5C.C7 mice express the transgenic TcR and respond to synthetic PCC peptide RADLIAYLKQATK *in vitro*. This mouse provides an excellent means to test PCC specific T cell responses both *in vitro* and *in vivo* as well as conduct adoptive transfer experiments. Adoptive transfer is a powerful method that allows the introduction of PCC reactive T cells into non-transgenic mice to study responses at varying T cell precursor frequencies.

Antigenicity of SAG:PCC peptide to 5C.C7 T cells

This experiment determines how potent the SAG:peptide conjugate is *in vitro*. It is a test of how well the antigen is taken up and presented by the APCs present in culture and whether the binding of SAG to MHC class II enhances presentation to T cells.

Lymph node T cells from adult 5C.C7 mice were incubated with varying amounts of either synthetic PCC peptide alone, SPEC-Y15A.C27S.N79C, PCC peptide and SPEC-Y15A.C27S.N79C.R181unconjugated or conjugated prior to addition in culture. MHC class II restricted T cell responses were measured by a 3-day ^{3}H thymidine incorporation assay. Methods used were standard techniques such as those described Current Protocols in Immunology (1998) Colligan, J., Kuisbeck, A.M. Shevach, E.M. and W. Strober eds. John Wiley & Sons, Inc (ref 25)

Results

Fig. 1 indicates that 5C.C7 T cells responded to 10,000 times less SAG:PCC conjugate than the peptide alone. Optimal response to the SAG:PCC conjugate occurred at 10pM compared to 100 nM for the same components added in unconjugated form. No response was observed to SAG: irrelevant peptide indicating that the response was specific to the PCC peptide.

Immunogenicity of SAG:PCC conjugate in 5C.C7 mice

This tests the ability of the SAG:peptide conjugate to generate an immune response *in vivo* and is a test of it's immunogenicity – that is to stimulate and expand peptide specific T cells.

(i) Adoptive transfer of 5C.C7 T cells into wild-type C57Bl/6 mice

Normal female C57Bl/6 recipient mice receive 5 x 10^6 5C.C7 lymph node cells IP 1 week prior to immunisation.

Immunisation protocol

Antigens were injected as a single subcutaneously (SC) dose as a stable emulsion with Freund's incomplete adjuvant in mature female C57Bl/6 mice that had previously received 5C.C7 T cells. Two mice were injected for each dose with one of:

1. PCC peptide alone (1 and 100 mg)
2. PCC peptide + SPEC-Y15A.C27S.N79C.R181
3. SPEC:PCC conjugate (20 ng)

Mice were sacrificed 10 days later and the draining mesenteric lymph nodes removed. 1 x 10^5 lymph node cells/well were cultured in duplicate with varying amounts of synthetic PCC peptide and the proliferative response of T cells measured by the 3 day ^3H thymidine incorporation assay.

Results

Figure 2 indicates that the lowest dose of SAG:PCC conjugate used to immunised 5C.C7 mouse was 20 ng and this produced optimal immunity equivalent to 100 mg of free PCC peptide. 1 mg of PCC peptide was non immunogenic. Thus the SAG:PCC conjugate was at least 10,000 times more immunogenic than free peptide. Irrelevant peptides coupled to SPEC generated no detectable immune response. It is likely that even lower doses of SAG:PCC conjugate will be immunogenic, increasing the effective difference in potency between conjugated and unconjugated PCC peptide to 100,000 times.

These studies show that SPEC-Y15A.C27S.N79C.R181acts as an efficient delivery vehicle for poorly immunogenic antigens such as synthetic peptides. Not only is the peptide significantly more antigenic *in vitro*, but this also translates into enhanced immunogenicity *in vivo*. The immunogenicity of the PCC peptide

increased by at least 10,000 times by coupling to the TcR binding defective superantigen SPEC-Y15A.C27S.N79C.

SPE-C mutant defective in MHC class II binding does not enhance antigenicity of the PCC peptide.

A recombinant mutant of SPE-C was created that disrupts the single zinc binding site to MHC class II. This mutant was coupled to synthetic PCC peptide and tested for its ability to stimulate 5C.C7 T cells *in vitro* compared to normal SPEC:PCC conjugate.

The results show that the mutant SPEC:PCC conjugate was no more antigenic than the SPEC + free peptide alone. This indicates that enhanced antigenicity is a result of SPE-C's ability to bind to cells expressing MHC class II, a function unique to superantigens.

Figure 3 shows data which reveals the importance of MHC class II binding to enhancement of antigenicity and that SPEC is not simply acting as a "non-specific" carrier protein.

Example 10: Coupling of multiple peptides

Coupling need not be limited to individual peptides. Because immune responses to peptides are tightly restricted by the MHC polymorphisms of the host, it might be appropriate in some circumstances, to immunise with sets of peptides to generate broad spectrum immunomodulatory agents. Multiple peptides representing various components of a larger antigen such as a virus, bacteria or other protein antigen may be coupled by procedures described above or modified versions therefore which would be clear to those skilled in the art, to provide a mixed peptide:SAG conjugate antigen response to increase the diversity of the conjugate. Moreover, the ratio of peptides could be easily controlled to fine tune the immune response to a more desired outcome.

In further embodiments of the present invention, and applying the principles described herein, the following can also be accomplished:

- MHC class I and class II restricted peptides may be combined to provide improved helper CD4 and cytolytic CD8 effector cells.
- Immunodominant peptides from more than one viral antigen may be combined to promote selective anti-viral immunity.

- Peptides from regions of viral antigens that do not normally predominate in the protective immune response but represent regions of the virus essential to its replication or life cycle and are by nature strongly conserved may be used. This is particularly important in developing vaccines against highly mutating viruses such as retroviruses (e.g. HIV).
- Peptides and other antigens can be combined together and delivered by the immunomodulators to enhance or modulate the immune response.

Example 11: Coupling of larger antigens and complex structures

Polypeptides and proteins can be coupled using the same procedures described above by reversible disulphide interchange to mutant SAGs. In addition, larger structures such as viruses can be "coated" with a TcR defective SAG by first treating the virus with a chemical that introduces a reactive sulphydryl group.

If the polypeptide has a naturally occurring exposed cysteine residues, coupling may be achieved to SAG directly without the need to introduce a reactive sulphydryl group. In this case, coupling would follow the established procedure outlined above.

Chemical coupling methods

If the polypeptide does not have a naturally occurring cysteine, there are two methods that introduced a reactive sulphydryl group

a. A cysteine residue can be introduced genetically into the recombinant peptide and the polypeptide expressed from a heterologous expression system (prokaryotic or eukaryotic)
b. A chemical coupling reagent can be employed to introduce a reactive sulphydryl into the target protein or larger structure. A number of chemicals can be employed to introduce reactive sulphur groups onto proteins and other structures. One such chemical is N-succinimidyl S-acetylthiolproprionate (SATA – Piece Chemicals) and its close analogue SATP. This chemical converts a free amino groups on a protein or larger structure to a protected sulphydryl group which is activated with hydroxylamine. This allows coupling of other sulphydryl containing proteins such as SPEC-Y15A.C27S.N79C.R181via a reducible disulphide bond.

Relevant techniques are described in Ref. 21, incorporated herein by reference.

Delivery of proteins known to generate protective immunity for a particular pathogen can be made more immunogenic by first conjugating the protein to a TcR ablated SAG. The polypeptide would be broken down internally by the APC to present multiple restricted peptide epitopes to the host immune system. Anti-viral immunity might be enhanced by adding on molecules that selectively target the virus to APCs such as dendritic cells.

Example 12: Multiple Sclerosis and EAE in mice

For multiple sclerosis, the predominant self antigen appears to be the Myelin Basic Protein (MBP) which is the major component of the myelin sheath. Experimental Allergic Encephalitis (EAE) is a well-established mouse model for the human disease multiple sclerosis. EAE can be generated by immunising susceptible mice with myelin basic protein (MBP) which produces anti-MBP reactive T cells that attack the myelin coating of nerves, leading to the encephalitic disease characterised by loss of motor control [18].

The EAE model can be used to examine the ability of mutant SAG:MBP peptides or mutant SAG:MBP protein conjugates to inhibit the start of the disease, or to suppress existing disease [19]. Peptides (both agonist and antagonist) from the myelin basic protein (MBP) will be tested for their ability to suppress the onset of the EAE disease in mice.

Example 13: Anti-viral responses and MHC class I restricted peptides

Mutant SAG:peptide conjugates could also serve to enhance MHC class I restricted CTL responses. CD8 positive CTL recognise peptides presented by MHC class I derived from viral infection and replication via the endogenous processing pathway. It has been shown however that there is significant cross-talk between the endogenous and exogenous pathway for peptides to be "shared" by both MHC class I and MHC class II molecules.

Protective cytolytic responses against viral infection or tumours are believed to require an obligate CD4 MHC class II dependent response as well as MHC class I restricted CD8 responses to provide long lasting protective immunity. Thus vaccines constructed from the conjugation of MHC class I restricted peptides and SAG mutants or a combination of both MHC class I and MHC class II restricted peptides would offer a flexible approach to designing efficient vaccines which promote both CD4 and CD8 responses.

The $LCMV_{33-41}$ peptide and the 318 transgenic mice

The 318 transgenic mouse is a C57BL/6 mouse with a transgenic TcR which recognises the lymphocyte choriomeningitis virus (LCMV) peptide in the context of the MHC class I antigen H-$2D^b$ [20]. The sequence of the active peptide is CKAVYNFATM which originates from the nucleocapsid protein. The 318 mouse will be used to model the ability of SPEC-Y15A.C27S.N79C.R181and other TcR defective SAGs to deliver MHC class I restricted peptides to CD8 cytotoxic T cells. Efficiency of delivery will be measured by the amount of SAG:LCMV conjugate required to generate a cytotoxic response against target cells pre-incubated with LCMV peptide (standard cytotoxic assay).

The ^{51}Cr release cytotoxicity assay to measure MHC class I restricted responses

Target cells (P814) are incubated with ^{51}Cr and pulsed with LCMV peptide for 1 hour at 37°C. Cells are washed by centrifugation and mixed with lymph node cells from immunise mice at varying E:T ratios.

Cells are centrifuged lightly and incubated at 37°C for 1 hour. Supernatant is removed and counted for ^{51}Cr to determine the degree of cell lysis.

In vitro *cytotoxic assays: JAM test*

Cytotoxic T lymphocytes (CTL) were generated by culturing lymph node cells from 318 mice for 5 days in 24 well plates ($2.5x10^6$/ml) in the presence of serially diluted sAg-peptide complexes or cysLCMV peptide. Cells were fed on day 3 with rhIL-2 and RPMI. At day 5, cells were washed twice and assayed for CTL activity by the JAM test (Matzinger, P., 1991, J immunol methods, 145:185-192). Briefly, EL4 cells were incubated with 5□Ci/ml [^{3}H]thymidine overnight, the next day LCMV peptide was added at 10nM for 3hrs. Cells were washed twice and used as target cells for LCMV-specific CTL generated with SAG-LCMV complexes as descibed above: CTL and EL4 cells were incubated together at various effector:target (E:T) ratios for 3 hrs. The percentage of specific lysis was calculated using the following equation:

$$\% \text{ Specific Lysis } \frac{S-E}{S} \times 100$$

S = retained DNA in the absence of killers (CPM), and E = retained DNA in presence of killers (CPM). Results are shown in Figure 6.

Synthetic LCMV peptide modified at position 8 (M8C) were coupled to SPEC-Y15A.C27S.N79C.R181using the same method as described above. SAG:LCMV was used to determine the *in vitro* response in lymph node cells from 318 mice.

Resistance to viral infection

Mice infected with LCMV succumb within 14 days to the cytopathic effects. Mice immunised against LCMV develop a CTL response which provides full protection against. Mice immunised with SAG:LCMV will be tested for their resistance to wild-type LCMV virus.

Example 14: Anti-tumour immunity- *In vivo* tumour protection.

Many novel cancer immunotherapies attempt to break host tumour tolerance by targeting potential tumour specific antigens (usually lineage specific or differentiation antigens) directly to dendritic cells. We have discovered that TcR defective SAG mutants usefully target tumour specific antigens to APCs and promote co-stimulatory signals that enhance antigen presentation. Initial studies employed a tumour model in the 318 TcR transgenic mouse.

Our initial experiments have shown that SAG-peptide complexes can inhbit tumor growth in the LCMV-tumor (the model was described by Dr. Ronchese and colleagues - *Hermans, I. et al,1997, Caner immunol Immunother 44:341-347)*:

Triple and quadruple TCR-/- SMEZ-2 were coupled to LCMV peptide and emulsified with an equal volume of incomplete Freund's adjuvant (IFA). SAg-LCMV was injected equivalent to 1µg peptide and compared with 100µg of peptide emulsified with IFA injected s.c. into the right flank of groups of C57BL6/J mice (n=5). Seven days after immunisation, mice were challenged with $1x10^6$ LL-LCMV tumour cells injected s.c. into the left flank. The LL-LCMV cells are Lewis lung carcinoma cells (LLTC) transfected with a minigene for the LCMV $_{33-41}$ peptide and can be recognized as targets *in vivo* in mice of H-2^b background. Mice were monitored every 3-4 days and sacrificed, once 1 mouse in each group had tumours reaching 16mm in diameter or 200mm^2 in area.

Tumor size is shown as the arithmetric means of the products of bisecting tumor diameters (Figures 7a and 7b show results of two separate assays)

The Lewis Lung carcinoma and the 318 transgenic mouse

A further model used for the study of anti-tumour immunity is a model of tumour protection using the 318 transgenic mouse. A Lewis Lung carcinoma cell line transfected with a gene expressing the LCMV glycoprotein provides a model to investigate the ability of 318 mice to reject tumours. This cell line has high metastatic potential.

Mice were immunised with SAG:LCMV peptide and then inoculated with tumour cells. The degree of metastatic foci were established at varying time points following inoculation and compared with non-immunised mice. Results similar to those described above were obtained.

Mice were also be inoculated with tumour cells and then immunised at varying time points following tumour inoculation to determine whether immunisation protects against established tumour growth.

Example 15: Increasing the antigenicity of a whole protein to T cells by coupling to SAG.

1 mg whole Pigeon Cytochrome C protein (PCC) (Sigma) was treated with 1 mg of the cross-linked reagent N-succinimidyl S-acetylthioproprionate (SATP)(Pierce) for 1 hour at room temperature at pH7.0. Excess cross-linker was removed by gel chromatography using well established protocols, and the PCC-SATP activated with 0.1M hydroxylamine and incubated with 100 μg recombinant SAG for 1 hour at pH8.5 to allow the proteins to couple. Conjugate was separated from free reactants by size exclusion chromatography according to well established protocols.

This method results in approximately 30% of the SAG forming a conjugate with PCC in a molar ratio of 1:1.

Conjugates were incubated with cultures of lymph node cells from T cells 5C-C7 mice and proliferation of T cells measured by ^{3}H thymidine incorporation after 3 days, according to well established protocols. Results of these studies are shown in Figure 5.

The results show a substantial increase in the antigenicity towards PCC protein when conjugated to either SPEC or SMEZ. They further emphasises the

importance of binding of the SAG to MHC class II to achieve increased antigenicity.

Another model used to assess immunogenicity of SAG-CytC complexes was the 5C.C7 transgenic T cells. These were labeled with 5-(and -6)-carboxyfluorescein diacetate succinimidylester (CFSE) according to Fazekas de st. Groth, B.,et al, 1999, Immunology and Cell Biology 77:530-538. 2-5x10^6 transgenic CFSE -labeled cells were transferred ip into congenic B10.A mice. The mice were immunized 3days later as follows: SAG-CytC complexes were injected sc. at various concentrations (already referred to earlier herein) emulsified in IFA and compared with free CytC peptide and free peptide mixed with SAG, also in IFA. Proliferation of transgenic CFSE-labeled cells was monitored 2-3days later via FACScan analysis. Similar results to those above were obtained.

Some of the advantages and features of the exemplary TcR defective immunomodulatory conjugates of the present invention are the following:

a. The SAG is totally defective in binding to all TcRs and thus will be non-toxic in vivo.
b. Coupling of peptides is simple, efficient and reversible and broadly applicable.
c. The SAG:peptide conjugate is soluble.
d. SAG binding to MHC class II enhances APC activation of immunogenic and non-immunogenic moieties.

Although the present invention has been described with reference to certain preferred embodiments it will be understood that variations, which are in keeping with the broad principles and the spirit of the invention, are also contemplated to be within its scope.

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What is claimed is:

1. Immunomodulator which comprises an antigen-presenting- cell (APC) targeting molecule coupled to an immunomodulatory antigen, wherein said APC-targeting molecule mimics a superantigen but does not include a fully functional T-cell receptor binding site.
2. Immunomodulator which comprises an antigen-presenting cell (APC) targeting molecule coupled to an immunomodulatory antigen, wherein said APC-targeting molecule is a molecule which is structurally a superantigen but for a disrupted T-cell receptor binding site such that the molecule has little or no ability to activate T-cells.
3. An immunomodulator according to claim 1 or claim 2, wherein the T-cell receptor binding site, or at least a part thereof, of the antigen-presenting-cell (APC) targeting molecule has been modified by substitution or addition.
4. An immunomodulator according to claim 1 or claim 2, wherein the T-cell binding site of the antigen-presenting cell (APC) targeting molecule has been deleted.
5. An immunomodulator according to any one of claims 1 to 3, wherein the antigen-presenting cell (APC) targeting molecule is derived from *Staphylococcus aureus* and/or *Streptococcus pyogenes*.
6. An immunomodulator according to claim 5, wherein antigen-presenting cell (APC) targeting molecule is derived from SPE-C, SMEZ and/or SEA.
7. An immunomodulator according to claim 6, wherein the antigen-presenting cell (APC) targeting molecule is designated SPEC-Y15A as herein defined.
8. An immunomodulator according to claim 6 or claim 7, wherein the antigen-presenting cell (APC) targeting molecule is designated SPEC-Y15A R181Q.
9. An immunomodulator according to any one of claims 6 to 8, wherein the antigen-presenting cell (APC) targeting molecule is designated SPEC-Y15A.C27S.N79C.R181Q
10. An immunomodulator according to any one of claims 1 to 9, wherein the antigen-presenting- cell (APC) targeting molecule is coupled reversibly to an immunomodulatory antigen.

11. An immunomodulator according to any one of claims 1 to 10, wherein the immunomodulatory antigen is a protein, a polypeptide and/or a peptide.
12. An immunomodulator according to any one of claims 1 to 10, wherein the immunomodulatory antigen is a nucleic acid.
13. An immunomodulator according to any one of claims 1 to 12, wherein the immunomodulatory antigen is non-immunogenic when not coupled to the antigen-presenting cell (APC) targeting molecule.
14. An immunomodulator according to claim any one of claims 4 or 10 to 13, wherein the antigen-presenting cell (APC) targeting molecule is SPEC (-20-90).
15. Pharmaceutical composition comprising an immunomodulator according to any one of claims 1 to 14 and a pharmaceutically acceptable carrier, adjuvant, excipient and/or solvent.
16. Vaccine comprising an immunomodulator according to any one of claims 1 to 14.
17. Method of therapeutic or prophylactic treatment of a disorder which requires the induction or stimulation of the immune system, comprising the administration to a subject requiring such treatment of an immunomodulator according to any one of claims 1 to 14, of a pharmaceutical composition according to claim 15 or of a vaccine according to claim 16.
18. A method according to claim 17, wherein the disorder is selected from the group consisting of bacterial, viral, fungal or parasitic infection, autoimmunity, allergy and/or pre-neoplastic or neoplastic transformation.
19. Use of an immunomodulator according to any one of claims 1 to 14 for the preparation of a medicament for the therapeutic or prophylactic treatment of a disorder which requires the induction or stimulation of the immune system.
20. Use according to claim 19, wherein the disorder is selected from the group consisting of bacterial, viral, fungal or parasitic infection, autoimmunity, allergy and/or pre-neoplastic or neoplastic transformation.
21. Method of preparing an immunomodulator comprising the steps of:

a introducing a modification and/or a deletion into the T-cell binding site of an antigen-presenting cell (APC) targeting molecule which is structurally a superantigen, and

b coupling thereto and immunomodulatory antigen.

22. A method according to claim 21, wherein the antigen-presenting cell (APC) targeting molecule is selected from the group of SPE-C, SMEZ and SEA.

23. A method according to claim 21 or claim 22, wherein the antigen-presenting cell (APC) targeting molecule is SPE-C Y15A R181Q

24. A method according to any one of claims 21 to 23, wherein the antigen-presenting cell (APC) targeting molecule is designated SPEC-Y15A.C27S.N79C.R181Q

25. A method according to claim 21 or claim 22, wherein the antigen-presenting cell (APC) targeting molecule is SPEC (-20-90).

26. Method of increasing antigenicity of a compound, comprising the coupling of said compound to an antigen-presenting-cell (APC) targeting molecule, wherein said APC-targeting molecule mimics a superantigen but does not include a fully functional T-cell receptor binding site.

27. A method according to claim 26, wherein said APC-targeting molecule is a molecule which is structurally a superantigen but for a disrupted T-cell receptor binding site such that the molecule has little or no ability to activate T-cells.

28. A method according to claim 26, wherein the T-cell receptor binding site, or at least a part thereof, of the antigen-presenting-cell (APC) targeting molecule has been modified by substitution or addition.

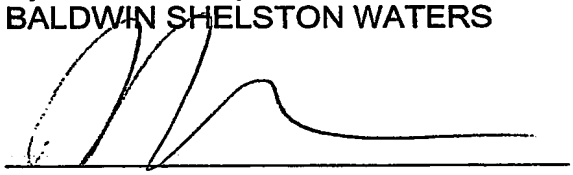
29. A method according to claim 26, wherein the T-cell binding site of the antigen-presenting cell (APC) targeting molecule has been deleted.

30. A method according to any one of claims 26 to 29, wherein the antigen-presenting cell (APC) targeting molecule is derived from *Staphylococcus aureus* and/or *Streptococcus pyogenes*.

31. A method according to claim 30, wherein antigen-presenting cell (APC) targeting molecule is derived from SPE-C, SMEZ and/or SEA.

32. A method according to claim 31, wherein the antigen-presenting cell (APC) targeting molecule is designated SPEC-Y15A as herein defined.
33. A method according to claim 31, wherein the antigen-presenting cell (APC) targeting molecule is designated SPEC-Y15A R181Q.
34. A method according to claim 31, wherein the antigen-presenting cell (APC) targeting molecule is designated SPEC-Y15A.C27S.N79C.R181Q
35. A method according to claim 31, wherein the antigen-presenting cell (APC) targeting molecule is SPEC (-20-90).
36. A method according to any one of claims 26 to 29, wherein the antigen-presenting- cell (APC) targeting molecule is coupled reversibly to said compound.
37. A method according to any one of claims 26 to 29, wherein the compound is selected from the group consisting of a protein, a polypeptide and/or a peptide, a carbohydrate or a nucleic acid.
38. A method according to any one of claims 26 to 29, wherein the compound is non-immunogenic when not coupled to the antigen-presenting cell (APC) targeting molecule.

AUCKLAND UNISERVICES LIMITED
By their Attorneys
BALDWIN SHELSTON WATERS



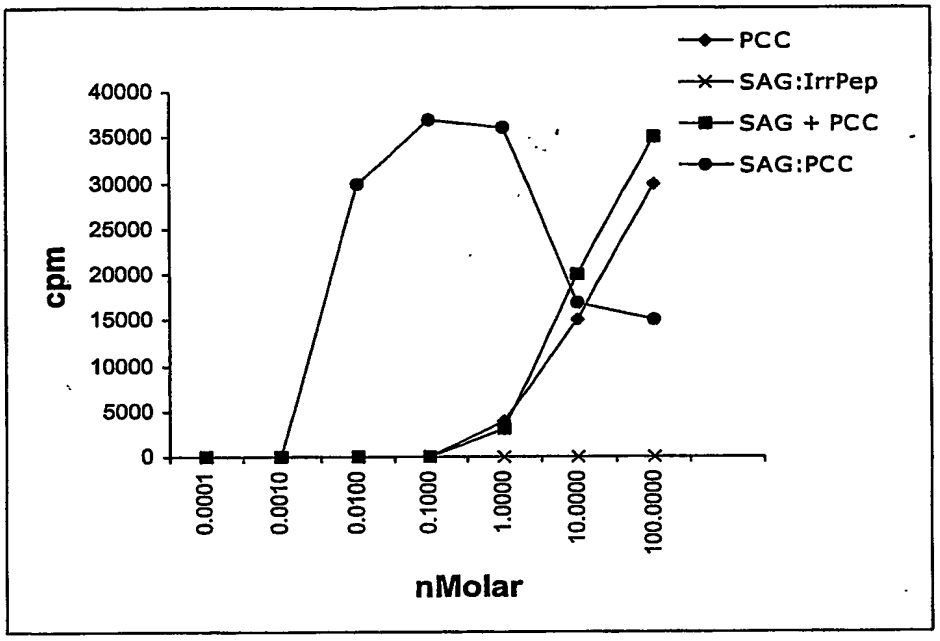

PCC
SAG:IrrPep
SAG + PCC
SAG:PCC
40000
35000
30000
25000
20000
15000
10000
5000
0
cpm
0.0001
0.0010
0.0100
0.1000
1.0000
10.0000
100.0000
nMolar

FIGURE 1

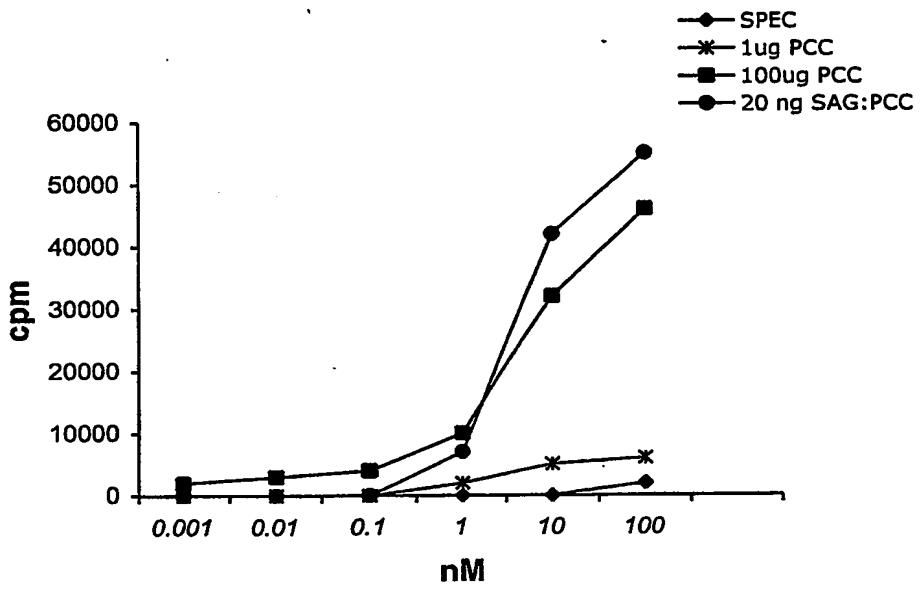

SPEC
1ug PCC
100ug PCC
20 ng SAG:PCC
60000
50000
40000
30000
20000
10000
0
cpm
0.001
0.01
0.1
1
10
100
nM

FIGURE 2

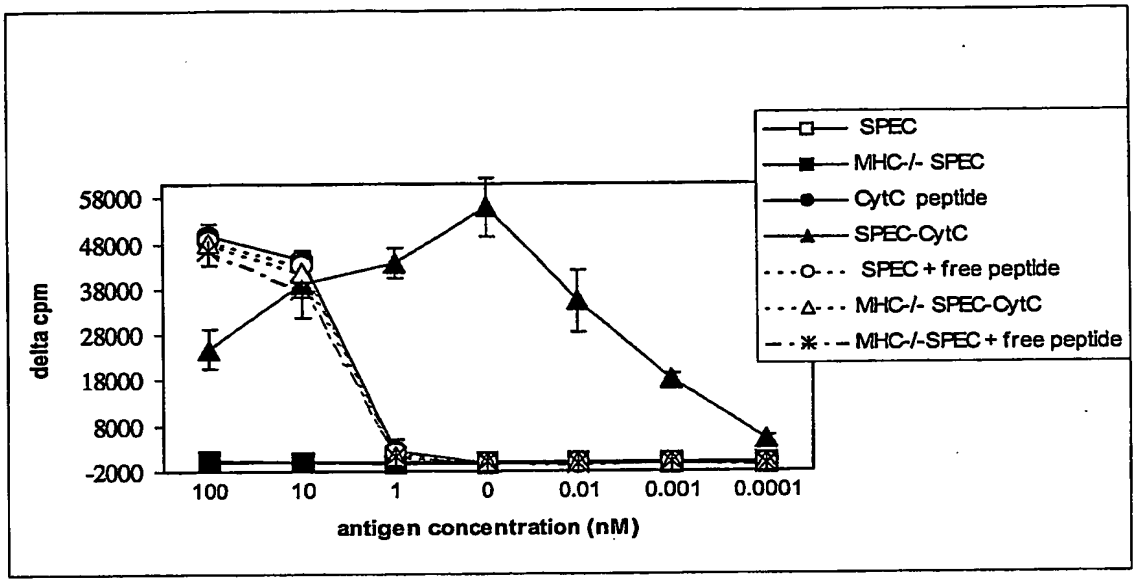

SPEC
MHC-/- SPEC
CytC peptide
SPEC-CytC
SPEC + free peptide
MHC-/- SPEC-CytC
MHC-/-SPEC + free peptide
delta cpm
58000
48000
38000
28000
18000
8000
-2000
100
10
1
0
0.01
0.001
0.0001
antigen concentration (nM)

FIGURE 3

SMEZ-2 mutants deficient in TCR binding

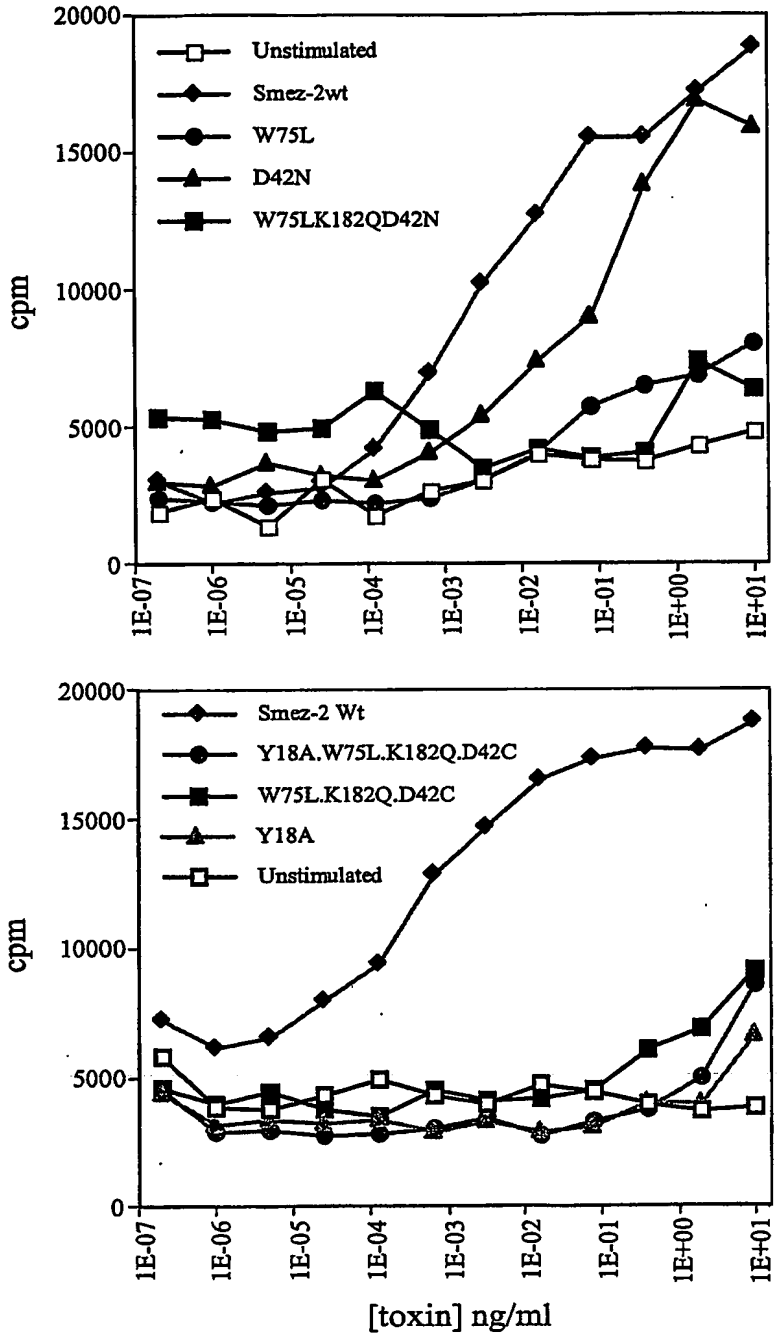
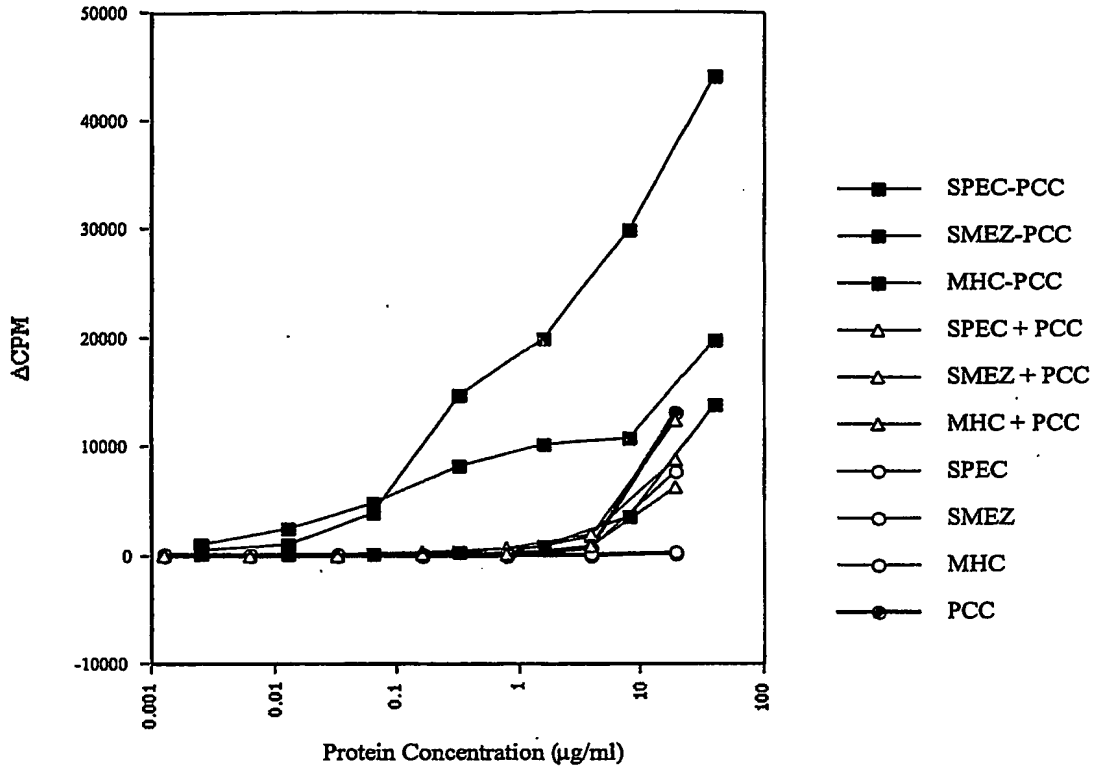


FIGURE 4

A

Proliferation Assay of 5C.C7 LN Cells with PCC-SAg Complexes



Coupling of whole PCC protein to SAg enhances protein immunogenicity *in vitro*

B

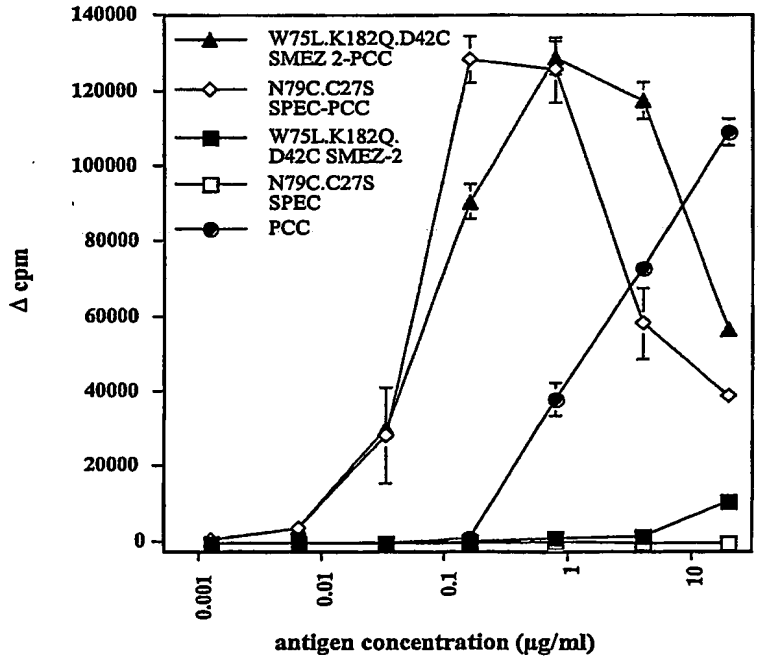


FIGURE 5

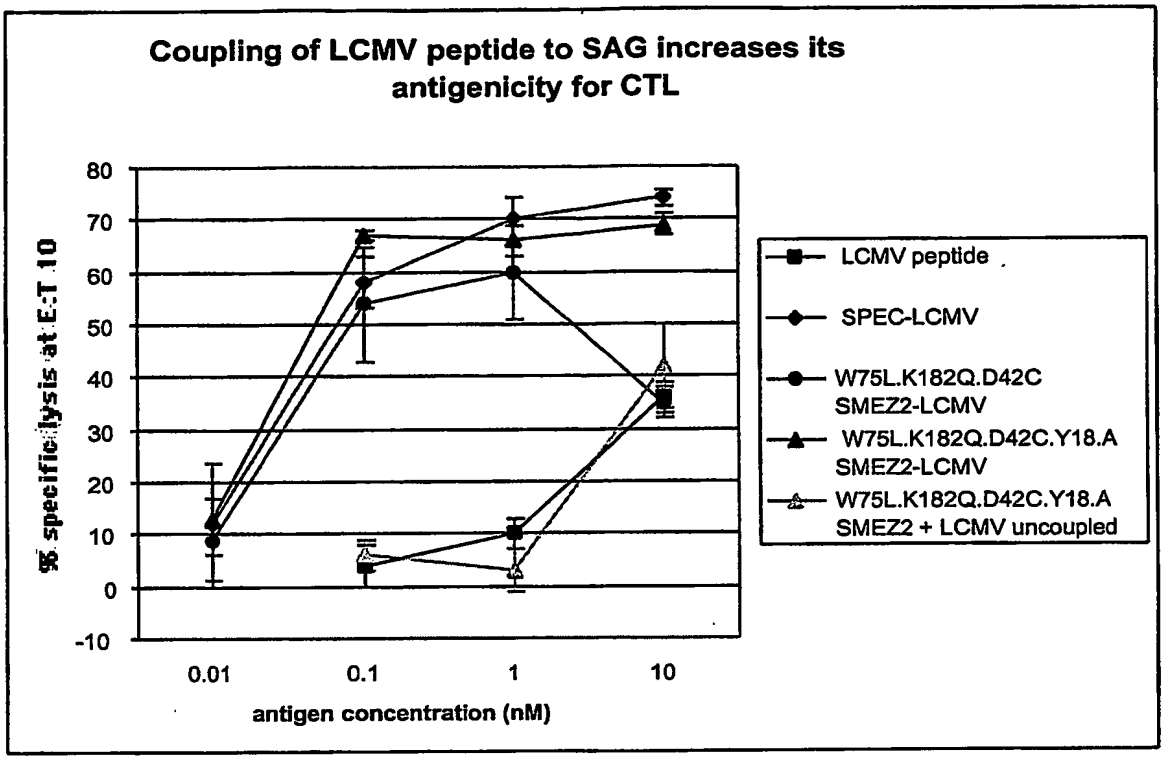

Coupling of LCMV peptide to SAG increases its antigenicity for CTL
% specific lysis at E:T 10
80
70
60
50
40
30
20
10
0
-10
0.01
0.1
1
10
antigen concentration (nM)
LCMV peptide
SPEC-LCMV
W75L.K182Q.D42C SMEZ2-LCMV
W75L.K182Q.D42C.Y18.A SMEZ2-LCMV
W75L.K182Q.D42C.Y18.A SMEZ2 + LCMV uncoupled

FIGURE 6

Inhibition of tumor growth by immunization with SMEZ-LCMV complexes

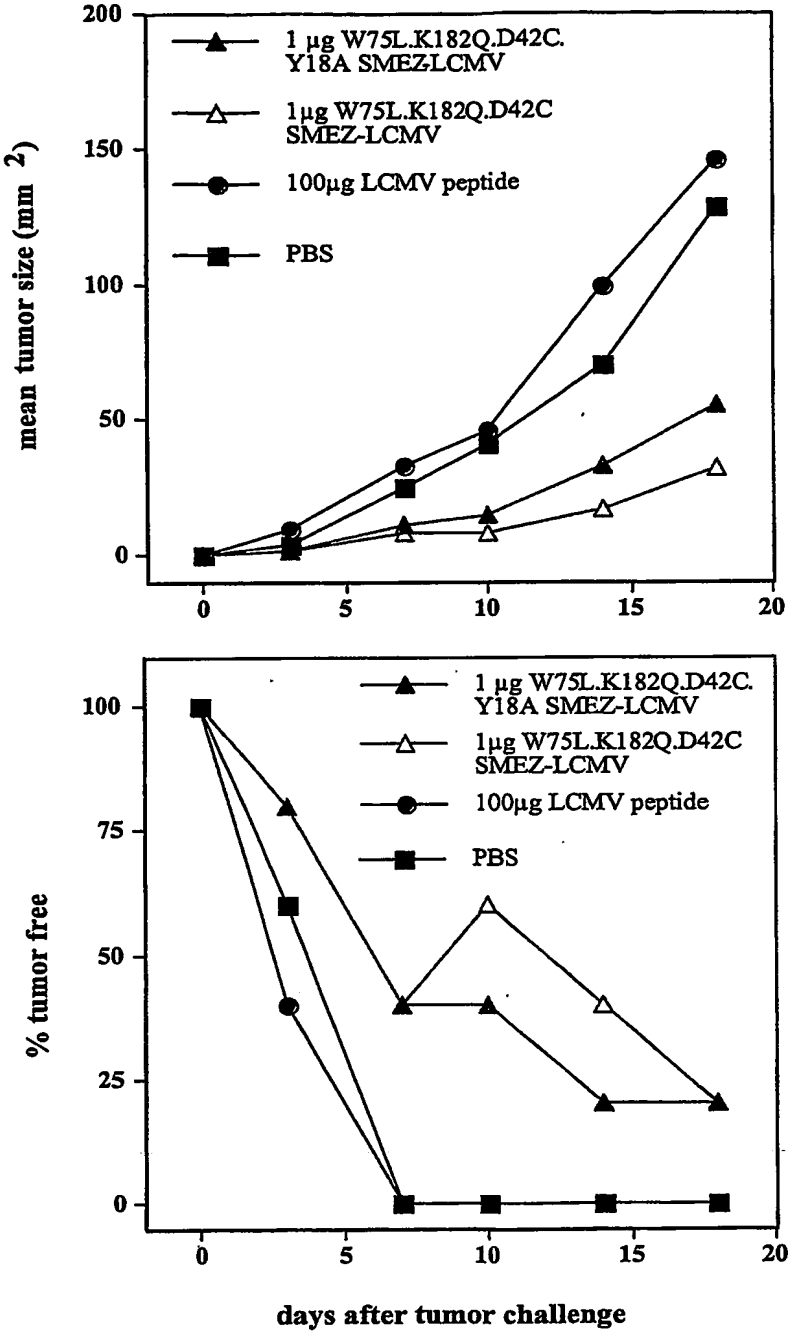


FIGURE 7a

Inhibition of tumor growth by immunization with SMEZ-LCMV complexes

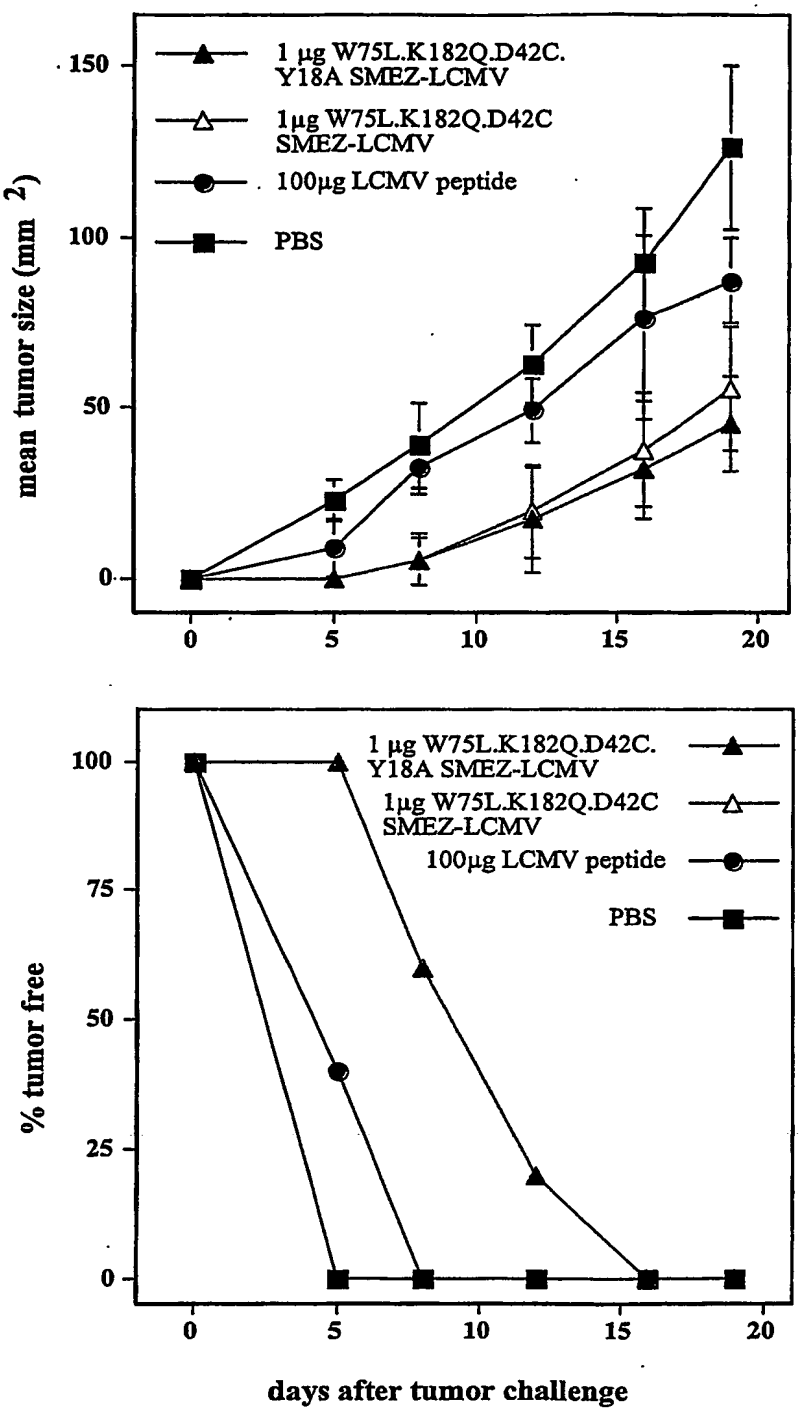


FIGURE 7b